



Mutational and biophysical robustness in a prestabilized monobody

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The fibronectin type III (FN3) monobody domain is a promising non-antibody scaffold, which features a less complex architecture than an antibody while maintaining analogous binding loops. We previously developed FN3Con, a hyperstable monobody derivative with diagnostic and therapeutic potential. Prestabilization of the scaffold mitigates the stability–function trade-off commonly associated with evolving a protein domain toward biological activity. Here, we aimed to examine if the FN3Con monobody could take on antibody-like binding to therapeutic targets, while retaining its extreme stability. We targeted the first of the Adnectin derivative of monobodies to reach clinical trials, which was engineered by directed evolution for binding to the therapeutic target VEGFR2; however, this function was gained at the expense of large losses in thermostability and increased oligomerization. In order to mitigate these losses, we grafted the binding loops from Adnectin-anti-VEGFR2 (CT-322) onto the prestabilized FN3Con scaffold to produce a domain that successfully bound with high affinity to the therapeutic target VEGFR2. This FN3Con-anti-VEGFR2 construct also maintains high thermostability, including remarkable long-term stability, retaining binding activity after 2 years of storage at 36 °C. Further investigations into buffer excipients doubled the presence of monomeric monobody in accelerated stability trials. These data suggest that loop grafting onto a prestabilized scaffold is a viable strategy for the development of monobody domains with desirable biophysical characteristics and that FN3Con is therefore well-suited to applications such as the evolution of multiple paratopes or shelf-stable diagnostics and therapeutics.

Developing a small, simple protein domain that includes a similar sized binding region to antibody complementarity determining regions (CDRs) is a successful strategy for

overcoming the complexity of antibody structure. Non-antibody scaffolds are single domains, typically smaller than 20 kDa in molecular weight, and mostly free of glycosylation or disulfide bonds that require eukaryotic expression (1). Critically, they exhibit comparable binding affinities to antibodies (2). Monobodies based on the fibronectin type 3 domain (FN3) are a popular scaffold for developing non-antibody therapeutics (2–5). The FN3 domain has an Ig-like fold and thus retains three of the CDR-like loops of an antibody variable fragment, but is structurally simple enough to be engineered for advanced non-antibody functions as the monobody scaffold (4–7). There are a number of unique monobody derivatives in active development including clinical Adnectins by LIB therapeutics (8) or ViiV Healthcare (9), the stability-enhanced Centyrins under ARO therapeutics (10, 11) or in a CAR-T format by Poseida Therapeutics (12), and also the related TN3 monobodies under Viela Bio (13).

An important consideration in the ability to evolve a non-antibody scaffold for binding is the combination of a high initial stability and a mutationally robust framework. The small size and lack of redundant framework regions in non-antibody scaffolds result in protein domains that will accumulate only a few mutations to their variable regions before stability becomes compromised (2, 14). Most monobody derivatives typically lose ~40 °C of thermostability upon evolution for binding (2), which often results in insoluble expression in bacteria that must be resolved through later rounds of evolution (15). Critical to the design of next-generation therapeutics, poor biophysical properties such as thermostability and poor or insoluble expression hinder scaffold “developability” and correlate to higher risk of failure in during clinical development (16, 17).

There are multiple approaches of improving the stability of protein folds (18). Protein stability exists between two critical thresholds, where a protein is able to fold into a stable, native three-dimensional shape but is also still dynamic enough to perform its functions (19, 20). As proteins evolve over time, the resulting mutations are more likely to be detrimental to

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stability of the native fold than neutral or positive and may reduce protein stability below the folding threshold (21). This is often considered as a natural trade-off between stability and activity and can be a severe limitation to directed evolution experiments (22, 23). Therefore, as a protein scaffold takes on mutations to a variable region to improve binding, it is at risk of deleterious losses in thermostability. Accordingly, proteins with improved initial thermostability are able to sample a larger proportion of this destabilizing sequence space as neutral mutations—they are more *mutationally robust*—which in turn increases the chance of reaching novel functions (24–27). In this way, prestabilization can enhance both evolvability—the ability of a protein to evolve new functions (23), and developability—the biophysical likelihood of successful development from a lead protein into a therapeutic drug (17).

From a drug development perspective, the native thermostability of a protein correlates with expression titre and improves critical quality attributes such as shelf-life (16), while also expanding the range of storage formulations which can be used in a drug product (28). Biological formulations need to remain stable for at least 2 years at 5 °C, and storage buffers are usually applied to reach this target. This has resulted in a standard set of buffer formulations across industry that are well-validated and focused on reducing aggregation or other loss of active protein (29). However, this sole focus on protein stability limits the exploration of nonstandard formulations, which benefit other critical quality attributes (28), such as controlling viscosity or osmolarity to lower the pain associated with injection (30–32).

Multiple, well-established techniques use phylogenetic information to derive thermostable homologs for a given protein family (18, 33–37). Here we have specifically applied consensus design, which identifies conserved residues within a protein family that are likely to be stabilizing (34, 37). Previously, we used consensus design to produce the prestabilized FN3Con monobody, which has substantially increased thermostability compared with similar fibronectin domains (38). The mutational robustness of FN3Con was then demonstrated by grafting binding loops from an FN3 scaffold that had been selected by laboratory evolution to bind lysozyme with high affinity (39). Whereas binding function was completely transferred, the trade-off in thermostability was negligible compared with that which occurred upon directed evolution of the original scaffold.

In the current study, we aimed to establish whether FN3Con can harbor valuable loop sequences that confer clinical inhibition of a target but were detrimental to stability in established scaffolds. For this study we chose an Adnectin domain that was previously subjected to directed evolution for high-affinity binding to the therapeutic target VEGFR2, with function gained at the expense of large losses in thermostability and increased oligomerization propensity (40, 41). We grafted binding loops from Adnectin-anti-VEGFR2 onto the FN3Con scaffold to produce a recombined domain that retained binding affinity. The FN3Con-anti-VEGFR2 graft was expressed in *E. coli* with little

aggregation and maintained characteristically high thermostability, including 24-month stability at 37 °C. An early exploration of buffer excipients produced further stability improvements. We discuss the implications of generating clinical leads by salvaging loop sequences from scaffolds with challenging biophysical features and the importance of designing highly “evolvable” constructs on downstream factors of scaffold “developability.”

Results

Transfer of affinity to a target by sequence grafting

We chose the Adnectin-anti-VEGFR2 monobody “CT-322” as a candidate for loop grafting to the hyperstable FN3Con in order to test our hypothesis that a stabilized scaffold can rescue stability losses accrued after evolutionary selection for high-affinity binding. This Adnectin was generated from mRNA display against a construct of seven extracellular domains of VEGFR2 fused with a human antibody Fc region, generating high-affinity binding ($K_D = 0.31$ nM) but with a 34 °C loss in thermostability (T_m) and also at cost to oligomerization resistance (40, 41).

Given the absence of structural information for the binding mechanism of Adnectin-anti-VEGFR2 to its large multidomain target, we used previously established loop sequence boundaries (39) to guide the transfer of evolved binding loops to FN3Con, designing FN3Con-anti-VEGFR2 [Fig. 1A and B]. Additionally, the entire Adnectin C-terminal tail was reported to be critical to high-affinity binding and was also transferred (40).

FN3Con-anti-VEGFR2 displayed high-affinity binding to VEGFR2 ($K_D = 0.72$ nM) [Table 1], very similar to the published affinity of Adnectin-anti-VEGFR2 ($K_D = 0.31$ nM) (41). We carried out affinity measurement through an orthogonal approach, where two independent methods provided a K_D range of 0.72 to 48.79 nM [Fig. 1, C and D and Table 1], with the K_D of 0.72 nM derived from Biacore data presenting the most robust fits to derive underlying equilibrium constants while controlling for confounding nonspecific binding and mass transport effects. The ELISA data validated this 2- to 3-fold difference in affinity between binders, although nonspecific binding likely increased the measured K_D for both monobodies.

Sequence-based grafting stabilizes an anti-VEGFR2 fibronectin

The Adnectin-anti-VEGFR2 undergoes irreversible thermal denaturation with a T_m of 50 °C \pm 0.4 °C, as measured by circular dichroism (CD), with visible precipitate upon cooling [Fig. 2A]. In striking contrast, FN3Con-anti-VEGFR2 unfolds reversibly with a T_m of 89 °C \pm 0.2 °C [Fig. 2B]. The T_m of the Adnectin-anti-VEGFR2 was not previously published (41), but our results closely match a precursor Adnectin-anti-VEGFR2 variant of similar affinity, thermostability, and loop sequences (40). This trajectory of loss in Adnectin thermostability presents the trade-offs that take place as affinity is further matured. In contrast, while the FN3Con scaffold loses \sim 10 °C of thermostability after loop grafting, scaffold stability remains

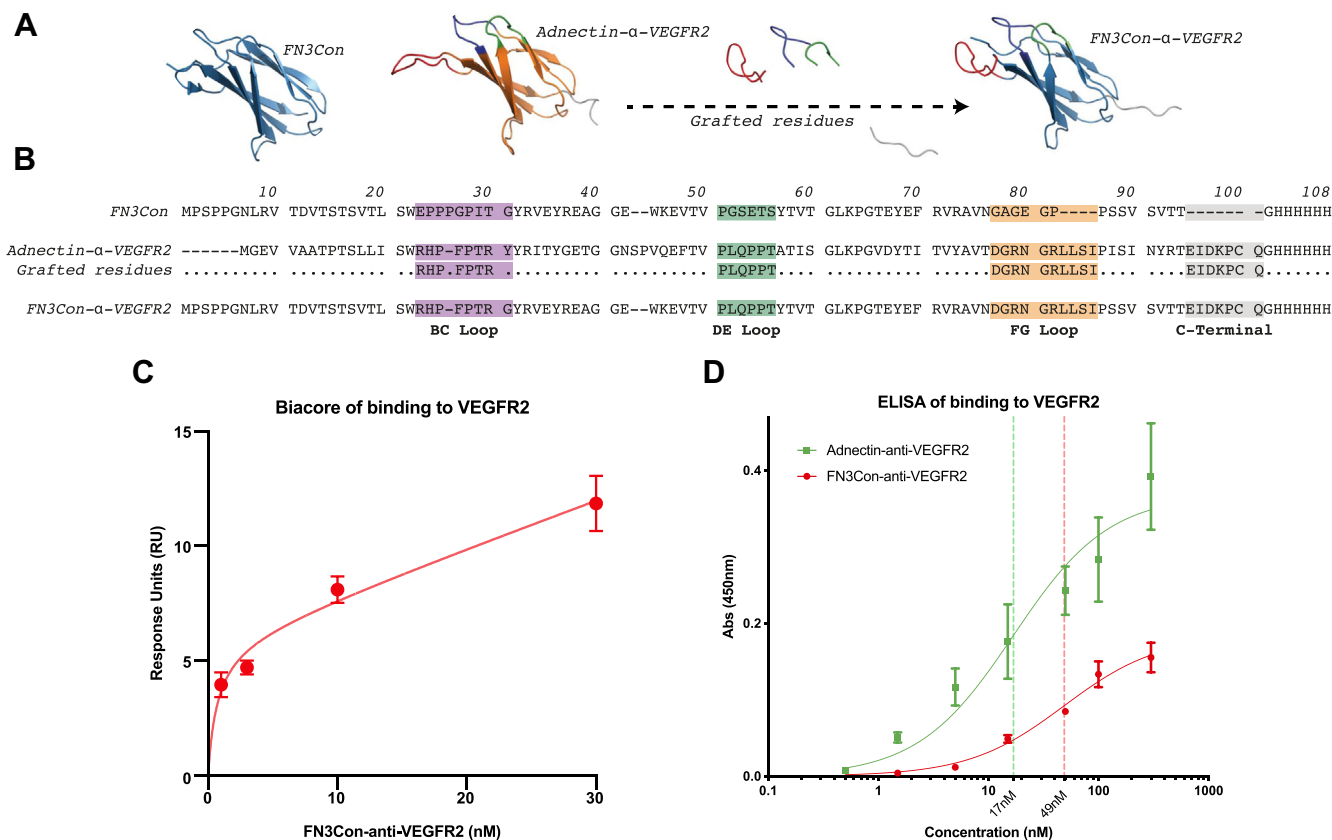


Figure 1. A and B, sequence-based grafting between monobody domains with loop sequences from (41). Affinity of FN3Con-anti-VEGFR2 to VEGFR2 was measured using (C) response units during steady-state of association in surface plasmon resonance (SPR), full data in Fig. S1, and (D) a general ELISA approach.

substantially higher than the parent Adnectin molecules, while also retaining reversible refolding.

FN3Con-anti-VEGFR2 retains stability and function at 36 °C for 2 years

We next investigated the effect of FN3Con-anti-VEGFR2 hyperstability on long-term stability (LTS) and binding activity, as a proxy for extended shelf-life. The Adnectin-anti-VEGFR2 sample completely aggregates within 1 month's storage at 36 °C in PBS [Fig. 2C]. In contrast, the FN3Con-anti-VEGFR2 remained in solution for at least 24 months, with ~30% of the total sample remaining as a monomer after 2 years storage at 36 °C [Fig. 2D]. Up to 50% of the FN3Con-anti-VEGFR2 high-order species observed formed between 0 and 6 months at 36 °C, after which oligomer formation

stabilized. Accordingly, FN3Con-anti-VEGFR2 presented extended LTS at 4 °C, remaining as a monomeric protein in extended trials up to 24 months of storage in PBS buffer [Fig. 2E]. Strikingly, high-affinity binding ($K_D \sim 157$ nM) to VEGFR2 was maintained after 24 months at 36 °C [Fig. 2F], although the observed affinity for the total sample is threefold lower than "fresh" FN3Con-anti-VEGFR2 ($K_D \sim 49$ nM). This suggests that only the ~30% monomeric fraction retains binding affinity to the target.

Matching protein thermal stability with formulation stability

Given the results from accelerated stability testing, our final investigation explored the effect of stabilizing excipients on further improving the shelf-life properties of the FN3Con-anti-VEGFR2 construct. After incubation at 40 °C for 30 days with

Table 1
Methodology and results for VEGFR2 binding experiments in Figure 1

Protein	Surface plasmon resonance (SPR) K_D	ELISA K_D
Adnectin-anti-VEGFR2	0.31 nM ^a	16.87 nM ± 4.26
FN3Con-anti-VEGFR2	0.72 nM ± 0.21	48.79 nM ± 6.78
Analysis		
Fold Difference	2.32	2.89
FN3Con-anti-VEGFR2 Model Fit	$R^2 = 0.95$	$R^2 = 0.98$
Immobilized protein	VEGFR2	VEGFR2
Measure	Change in refractive index	Concentration of Biotinylated protein

^a SPR K_D from (41), ELISA values are $K_D \pm SE$.

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five different buffer excipients (16), size-exclusion chromatography revealed that amino-acid excipients arginine, histidine, glycine, and aspartic acid produced a doubling of monomeric FN3Con-anti-VEGFR2 sample [Fig. 3]. Excipients such as Tween80 provide resistance to factors such as hydrophobic unfolding from shaking during storage (28, 42), and was added with no significantly greater detrimental effect on accelerated thermal stability than with PBS buffer alone.

Discussion

Previously, we have shown that FN3Con is robust to the stability–function trade-off (39). The data we present here is consistent with these findings, confirming our hypothesis that a stabilized scaffold can rescue functional regions from the stability losses accrued after evolutionary selection for high-affinity binding. Furthermore, our data confirms that grafting onto the FN3Con scaffold improves aggregation resistance over long-term storage. Importantly, we show that the stability of FN3Con is suitable for loop grafting to generate high-affinity binders against a clinically relevant target.

The Adnectin-anti-VEGFR2 monobody “CT-322” was the first of the Adnectin derivative of monobodies to reach clinical trials (41). Adnectin-anti-VEGFR2 showed potent efficacy with an antiangiogenic effect in preclinical models of pancreatic cancer (43), colorectal carcinoma and glioblastoma (41, 44), brain tumors (45), and Ewing’s sarcoma (46). Phase 1 clinical studies displayed clinical safety and acceptable pharmacokinetics to support phase 2 studies (47), although only some patients presented stable disease (48).

Unfortunately in phase 2 glioblastoma trials, the Adnectin derivative did not produce the required efficacy for continuation of studies (48, 49). This was suspected to be caused by a loss of inhibition effect during translation of this drug from the preclinical setting to human trials (49), which could result from pharmacokinetic issues with the Adnectin scaffold. Related to this could be the emergence of anti-Adnectin antibodies in 31 of 38 patients in the phase 1 study (47, 48). If this failure of clinical translation is driven by issues of developability, such as oligomerization, immunogenicity, and pharmacokinetics, then protein stabilization may salvage a potentially valuable clinical inhibitor.

An underlying explanation for the biophysical limitations of Adnectin-anti-VEGFR2 is that the evolution for high-affinity binding came at a critical cost to stability (T_m decrease of 30 °C) and oligomerization resistance [Fig. 2C] (41). The evolution of high-affinity VEGFR2 binding in the Adnectin scaffold resulted in a step-wise trade-off of thermostability for binding over a range of precursor loop sequences [Fig. 4A] (40, 50), including the loss of reversible refolding evident in the parent FNfn10 scaffold (38, 51). Despite its clinical failure, recent studies have sought to improve the Adnectin’s pharmacokinetic properties by increasing monomer size with a proline–alanine–serine repeat sequence (PAS-ylation) (52). Our loop grafting study examined the ability of FN3Con to mimic the targeted binding of antibodies, supporting the assertion that

consensus design of FN3Con resulted in a fibronectin domain that is robust to the mutational load of evolution for function.

To design FN3Con-anti-VEGFR2, we used available sequence information from the generation of Adnectin-anti-VEGFR2. There is a historic body of work related to rationally grafting CDR loops between antibodies (53). In practice, this grafting technique involves a balance of transferring residues involved in binding and removing stabilizing residues in the accepting scaffold (18, 54), which may reveal certain pairs of donor and recipient scaffolds to be graft-incompatible (39). The Adnectin-anti-VEGFR2 binding loops were compatible with the FN3Con scaffold, given the almost-complete transfer of affinity. This supports the approach that loop grafting to a more stable scaffold by sequence alone can be an effective strategy for the initial transfer of affinity between fibronectin-like domains, followed by focused rounds of redesign (39, 55).

Furthermore, there is growing evidence that monobody scaffolds can take on valuable loop sequences and their resulting affinity to clinical targets (56), such as in the transfer of anti-HER2 CDRs (57) or the anchoring of peptides (58). However, it is unclear how much of the previously published antibody CDR loop sequences can be transferred to monobody domains, especially if it is a complicated process that involves iterative redesign (55). Additionally, if there are already established antibodies in the clinic, then there may be little need for an antibody mimic, unless they can provide meaningful advantages such as short half-life for radiolabeled imaging (5, 59, 60). Nevertheless, it may be more efficient instead to evolve new binding loops *de novo* with display technologies (61, 62). If the FN3Con scaffold is hyperstable against destructive anti-VEGFR2 binding loops, it may be robust to directed evolution for aspects such as even greater affinity or the addition of a second binding surface.

In terms of biophysical properties generally considered under the concept of “developability” (16), FN3Con-anti-VEGFR2 initially presented improved features over the Adnectin in terms of thermal stability and high-yield, soluble bacterial expression (not shown). The improved thermal stability of this construct then led to favorable features of accelerated stability (AS) [Fig. 4B]. Although it is unclear whether higher-order species in the 2 years AS sample occurred due to self-association or to disulfide bonding between monomers as BME became inactive (63), the reduction in binding affinity to VEGFR2 matched the loss of monomer in the sample. As a result, in this investigation we see the flow-on effects from the focus on thermostability alone to a broader set of biophysical features and a link between the concepts of evolvability (27, 64) and developability (16).

Similar to our protein design approach, the purposes of excipient design for stabilization are to either reduce deviation from the native protein conformation or protect that state from collision with other molecules (65, 66). As there is a body of evidence around the interaction of salts negatively destabilizing FN3 proteins (67), we explored excipients that complement the enhanced protein stability of FN3Con.

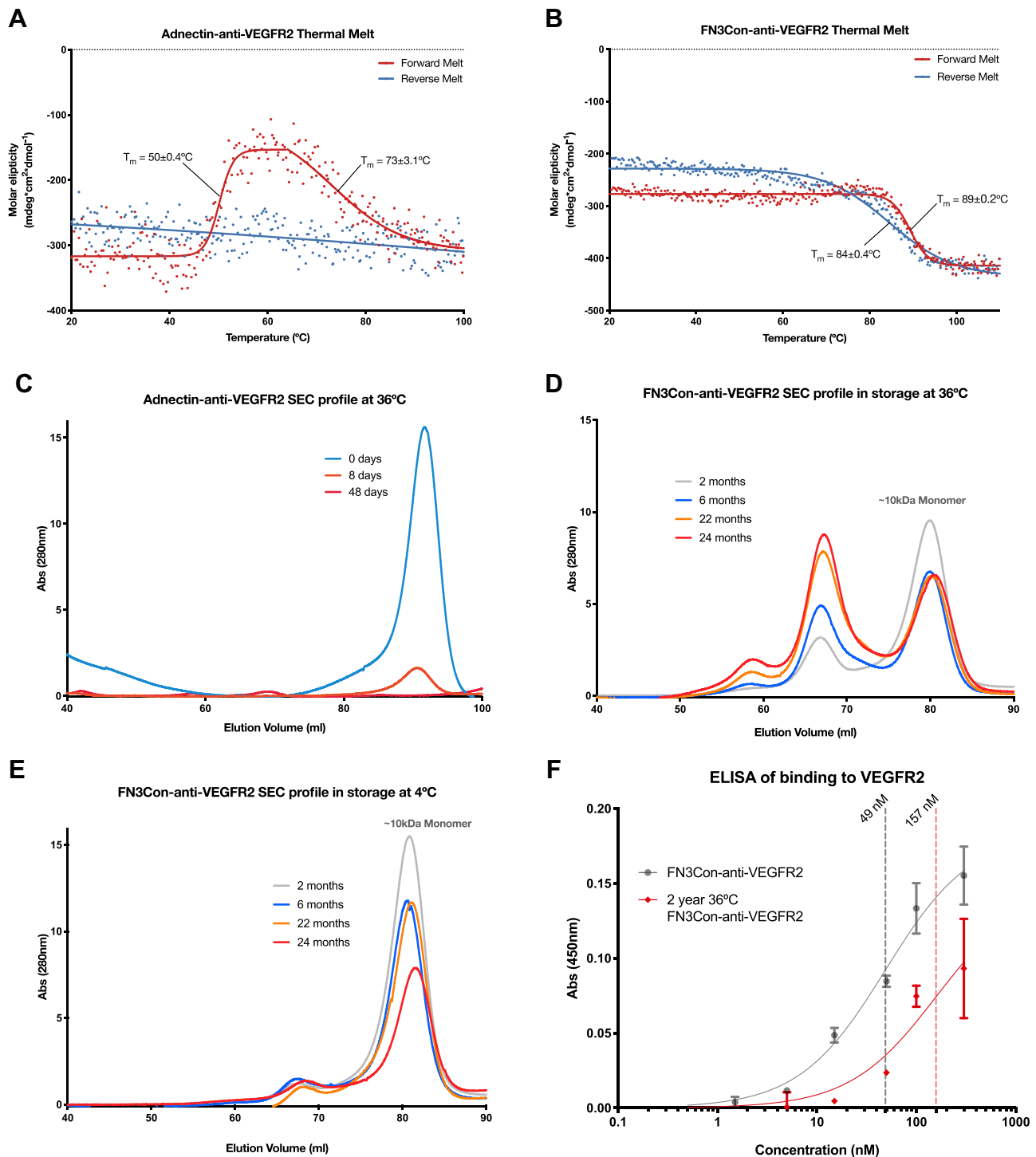


Figure 2. Circular dichroism (CD) thermal melts of (A) Adnectin-anti-VEGFR2 (T_m of $50\text{ }^\circ\text{C} \pm 0.4\text{ }^\circ\text{C}$) and (B) FN3Con-anti-VEGFR2 (T_m of $89\text{ }^\circ\text{C} \pm 0.2\text{ }^\circ\text{C}$), a reverse melt (blue) indicates reversible refolding in FN3Con-anti-VEGFR2. C, the Adnectin-anti-VEGFR2 sample aggregates completely over 1 month at $36\text{ }^\circ\text{C}$. D, more than 1/3 of FN3Con-anti-VEGFR2 sample is still present as a monomer after 2 years of $36\text{ }^\circ\text{C}$ storage, with the remaining 2/3 present as higher-order oligomers. E, the graft also shows limited aggregation during $4\text{ }^\circ\text{C}$ storage. F, ELISA of the $36\text{ }^\circ\text{C}$ 24-month sample shows a matching threefold loss in affinity (K_D of 49–157 nM).

In this study, accelerated stability trials confirmed the protective effect of amino acid excipients (29), as the addition of arginine, glycine, aspartic acid, and histidine doubled the amount of monomeric FN3Con-anti-VEGFR2 in accelerated conditions. The dominant stability improvement of

arginine over glycine [Fig. 3] could be due to the complex interaction of the arginine guanidinium group (68), which suppresses protein unfolding by increasing the energy barrier between folded native-state and aggregation-prone intermediates (69, 70). This can be seen as a further extension

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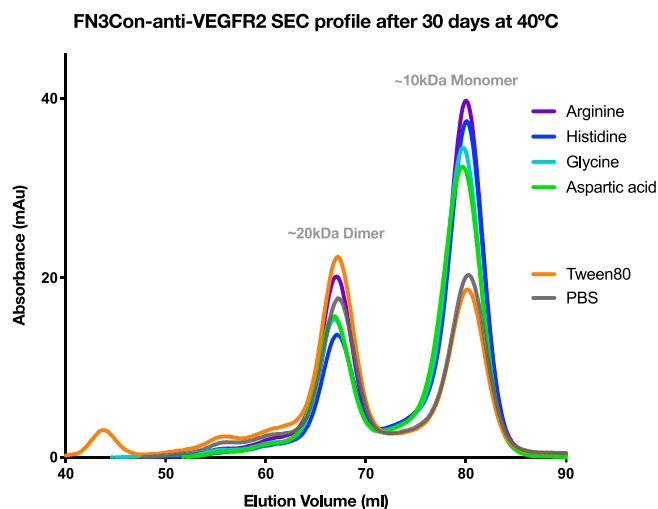


Figure 3. SEC elution profiles of FN3Con-anti-VEGFR2 after accelerated stability trials with a range of buffer excipients. All amino acid buffers slowed the loss of monomer. However, dimers appear at a similar rate between all samples, which could be linked to the generation of disulfide-bonded dimers due to degradation of BME. Tween80 was not significantly more destabilizing to the monomer than PBS buffer alone.

of the consensus protein design approach, which initially “smoothed out” this folding landscape and slowed the rate of unfolding in the native FN3Con conformation (38, 71). Histidine also provided improved resistance to thermal aggregation, and its effect is well-explored in industry formulations (72).

Generating highly stable biological scaffolds provides greater freedom to use nonstandard buffer formulations. As biological therapeutics must display a 5 °C shelf-life of up to 2 years, storage buffers are often designed with an aim of protection from aggregation or chemical modification of the protein (28). With hyperstability built into a protein scaffold, formulation scientists can instead focus on excipients that optimize function (66), such as for particulate drug delivery depots that overcome the short pharmacokinetic half-life of these small protein scaffolds, for freeze-dried and aerosolized suspensions, or in antimicrobial preservatives for long-term shelf-stable formulations.

These formulation experiments are important for designing differentiated applications from the dominant antibody scaffold, where the FN3Con domain may instead find its place in niche applications (5). The improved thermostability of the FN3Con monobody could allow for greater control over self-association intermediates in order to create novel drug modalities (5). Furthermore, our demonstration of stability and activity over 1 to 2 years supports the use of consensus-designed monobodies in diagnostic settings that cannot utilize cold-chain distribution, such as field applications of microfluidic devices (73).

Overall, the results of this study show the flow-on effects of prestabilizing a protein scaffold, from a greater robustness to evolutionary mutations, improved biophysical properties for clinical development, and potentially enabling a greater range of biomedical applications.

Conclusions

The initial aims of this study were to examine whether the engineering for hyperstability in FN3Con allowed for improved “evolvability” of clinical function in this hyperstable monobody derivative compared with other scaffolds that are progressing through the clinic. The complete transfer of affinity to a target was successful through sequence-based grafting alone, and the 40 °C increase in thermostability between monobody scaffolds suggests that the FN3Con scaffold is amenable to taking on affinity to a therapeutic target. Further to these aims, the connection of high thermal stability to biophysical aspects of “developability” was investigated. LTS trials show an active monomeric fraction after 2 years at 36 °C, and accelerated stability trials in a range of standard excipients show that there is much room for improvement in extension of that shelf-life. These are promising results for continuing development of the FN3Con scaffold. In future, critical investigations will need to utilize these advanced biophysical features to create novel constructs that provide differentiated patient benefit such as the evolution of multiple paratopes or in shelf-stable applications.

Experimental procedures

Protein expression and purification

Genes encoding Adnectin-anti-VEGFR2 and FN3Con-anti-VEGFR2 were chemically synthesized and provided in a pD444-CH (C-terminal 6x His tag, ampicillin resistance) vector by DNA2.0. The resulting plasmids were transformed into competent C41 *E. coli* cells for expression. A single colony from each transformation was picked and grown overnight at 37 °C in 100 ml of 2xYT (16.0 g/l tryptone, 10.0 g/l yeast extract, 5.0 g/l NaCl) media containing 100 µg/ml of ampicillin. These cultures were then used to seed 1 l of 2xYT media. Cultures were induced at an OD600 of 0.9 with IPTG (0.5 mM final concentration) and grown for a further 4 h at 37 °C. The cells were harvested by centrifugation. Adnectin-anti-VEGFR2 and FN3Con-anti-VEGFR2 had their cell pellets resuspended in 5 ml/g of native lysis buffer (50 mM NaH₂PO₄, 300 mM NaCl, 10 mM imidazole, pH 8.0) and were lysed by sonication. Cell debris was removed by centrifugation and incubated in lysis buffer +40 mM beta-mercaptoethanol to reduce disulfide bonds. Recombinant protein was then isolated from the supernatant by nickel affinity chromatography using loose Ni-NTA resin (Sigma). Protein eluted from Ni-NTA resin was filtered and then loaded onto a size-exclusion column (Superdex 75 16/60, GE Healthcare) equilibrated in PBS (140 mM NaCl, 2.7 mM KCl, 10 mM PO₄³⁻, 4 mM beta-mercaptoethanol pH 7.4) for biophysical characterization. Protein concentration was determined by Nanodrop ND-1000 (Thermo Fisher), and protein was stored at 4 °C until use.

Biotin conjugation of proteins

Biotin was conjugated to lysines, which are on the nonbinding loops of FN3Con-anti-VEGFR2 and Adnectin-

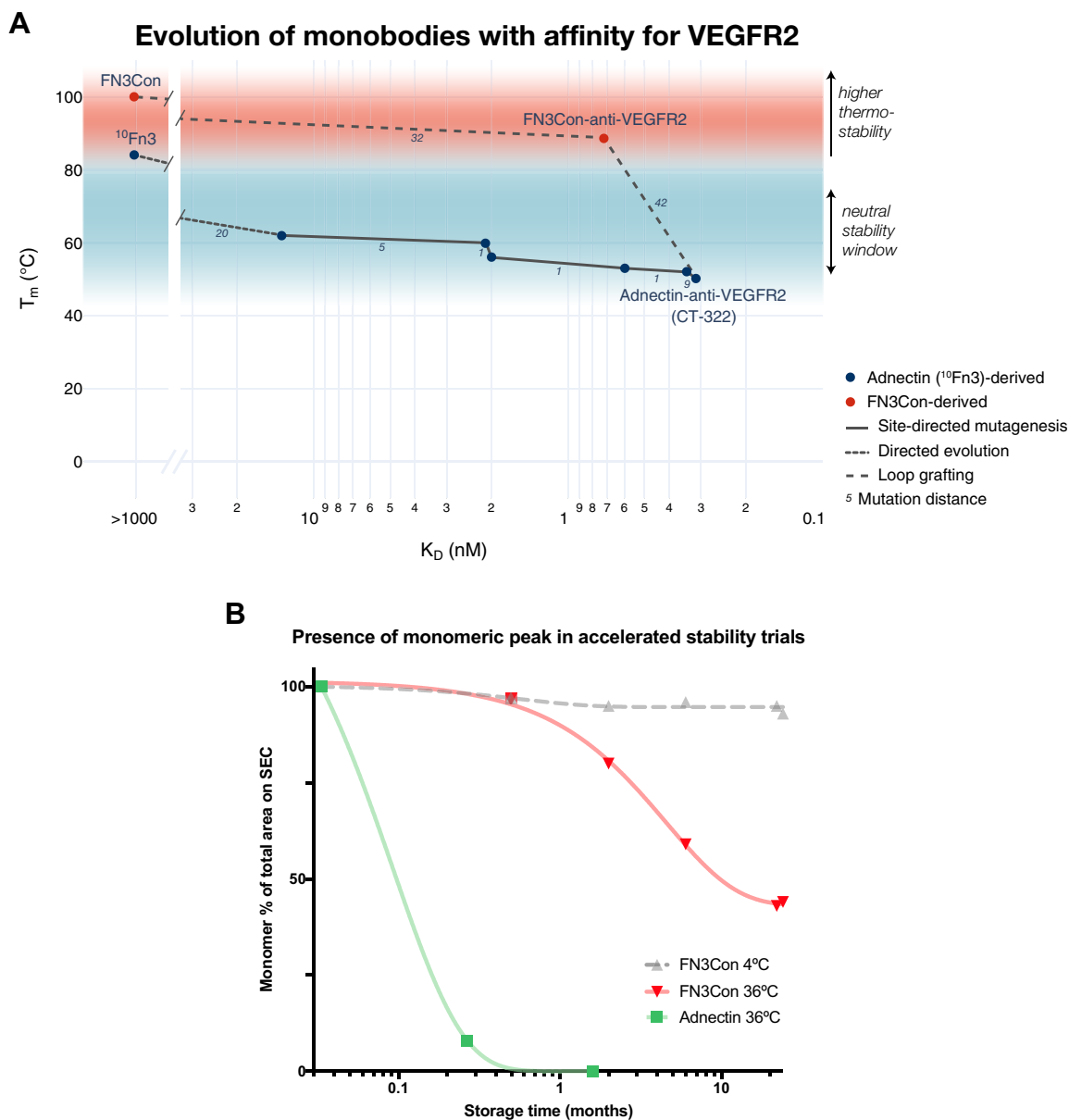


Figure 4. A, change in protein T_m as loops of the Adnectin scaffold were evolved for stronger binding to VEGFR2 (40, 41), transfer of binding loops to FN3Con retained similar affinity while heavily improving thermostability. Data in Table S1. **B, this improvement in thermal stability produced a related increase in LTS under elevated temperatures.** The Adnectin aggregated with no stable oligomers, so later monomer percentages were compared against the day 1 values (green). As FN3Con remained at ~ 1 mg/ml over the course of each trial, monomer percentages were calculated from the total peak area within each timepoint (red, gray).

anti-VEGFR2, to improve sensitivity and loading in ELISA or BLItz binding assays (EZ-Link Sulfo-NHS-LC-Biotinylation Kit, Thermo Fisher 21435).

Binding studies

SPR

The binding affinity of FN3Con-anti-VEGFR2 was measured using surface plasmon resonance with a 30 μ l/min flow rate at 25 $^{\circ}$ C (BIAcore T-100, GE Healthcare). VEGFR2 domains (Sino Biological, 10012-H08H) were conjugated on a CM5 sensor chip through NHS/EDC activation and ethanolamine deactivation. HBS-EP (10 mM HEPES, 150 mM NaCl, 0.005% (v/v) Tween 20, 0.1% BSA pH 7.4)

was used as the running buffer, and FN3Con-anti-VEGFR2 was prepared in serial dilutions of HBS-EP. Multicycle kinetics were performed with 240 s association of FN3Con-anti-VEGFR2 and 800 s dissociation with blank HBS-EP buffer, followed by regeneration of the surface (30 s Glycine-HCl pH 1.5 followed by a 150 s stabilization period, then 30 s NaOH pH 10 followed by a 300 s stabilization period).

ELISA

96-well plates were coated overnight with 10 μ g/ml VEGFR2, shaking at 4 $^{\circ}$ C. Wells were washed with PBS and blocked with milk blocking solution (5% powdered milk in

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PBS-T) for 2 h. Wells were washed again and incubated with biotinylated anti-VEGFR2 proteins for 1 h before washing and incubation with antibiotin HRP solution for another hour. After a final wash, TMB was added to wells and left to incubate for up to 30 min. Fluorescence at 415 to 450 nm was read by a plate reader for each well.

Binding calculations

High nonspecific binding was a limiting factor in the analysis of affinity especially when attempting to determine kinetic constants that were compromised also by a large component of mass transport during association and ligand rebinding during dissociation. Consequently, the steady-state value for association of each FN3Con-anti-VEGFR2 concentration was determined at 190 s after injection, and a K_D was calculated using a plot of RU values at this point as a function of concentration with GraphPad Prism and a simple Langmuir type fit ($Y = B_{max} * X / (K_d + X) + NS * X + Background$, where Y is the RU, X is the concentration in nM, and Background is constrained to 0).

The ELISA data was also limited by nonspecific binding; however, the aim of this experiment was to provide a comparison between Adnectin-anti-VEGFR2 and FN3Con-anti-VEGFR2. ELISA data was plotted in GraphPad Prism, and a one-site binding (hyperbola) model ($Y = (B_{max} * X) / (K_d + X)$) was applied to calculate the K_D as the midpoint of signal increase, this was presented as K_D alongside the R^2 measure of fit. The Biacore analysis was presented as the most representative estimation of affinity, as the steady-state approach allowed for better understanding of the effect of nonspecific binding.

Thermal stability

Thermal stability of purified Adnectin-anti-VEGFR2 and FN3Con-anti-VEGFR2 was measured by circular dichroism (CD). CD measurements were performed using a Jasco 815 spectropolarimeter with 0.2 mg/ml protein in PBS used in a 0.1 cm path-length quartz cuvette. Thermal denaturation was measured by observing signal changes at 222 nm during heating at a rate of 1 °C/min. The melting temperature (T_m) was obtained by fitting to a sigmoidal dose–response (variable slope) equation ($Y = Bottom + (Top - Bottom) / (1 + \exp((V50 - X) / Slope))$).

Long-term stability (LTS)

LTS was measured by diluting purified protein to 1 mg/ml in PBS + 2 mM beta-mercaptoethanol pH 7.4, then storing in 1.7 ml Eppendorf tube in a temperature-controlled rooms at 4 °C, 21 °C and 36 °C. Stability was investigated as the size of monomer peak on a size-exclusion column (Superdex 75 16/60, GE Healthcare) after injection of 100 µl volumes, 0.2 µm filtered, from each temperature fraction at 3, 6, 12, 22, and 24 months of storage. Peak areas were integrated by GE Unicorn software and percentages calculated from individual peak area divided by total area under the curve.

Accelerated stability

Accelerated stability was measured by diluting purified protein to 1 mg/ml in 2 mM Beta-mercaptoethanol pH 7.4 with 62.4 mM of glycine, histidine, aspartic Acid, arginine amino acids, or Tween80, then storing in 1.7 ml Eppendorf tube in a heat block at 40 °C for 30 days. Stability was investigated as the size of monomer peak on a size-exclusion column (Superdex 75 16/60, GE Healthcare) from injection of 500 µl volumes, 0.2 µm filtered, from each sample.

Data availability

All data are contained within the article.

Supporting information—This article contains [supporting information](#) (40, 41).

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Author contributions—P. G. C., B. T. P., and A. M. B. designed the study and drafted the paper. P. G. C. and L. L. T. performed data acquisition and interpretation. J. S. G., B. T. R., S. S. B., D. E. H., and M. B. assisted in data analysis, acquisition, and interpretation. D. E. H., R. J. F., T. P. M., C. J. J., and A. M. B. contributed to writing and conceptual advances.

Conflict of interest—The authors declare no competing financial interests.

Abbreviations—The abbreviations used are: CDR, complementarity determining region; FN3, fibronectin type III; LTS, long-term stability.

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